Thiols, Malonaldehyde and Total Antioxidant Status in the Turkish Patients with Type 2 Diabetes Mellitus

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DUMAN, B.S., ÖZTÜRK, M., YILMAZER, S. and HATEMİ, H. Thiols, Malonaldehyde and Total Antioxidant Status in the Turkish Patients with Type 2 Diabetes Mellitus. Tohoku J. Exp. Med., 2003, 201 (3), 147-155 — Non-insulin-dependent (Type 2) diabetes mellitus (NIDDM) is a risk factor for cardiovascular diseases (CVD). Oxidative stress mechanisms are often reported to be implied in type 2 diabetes mellitus. In order to determine their clinical relevance, we investigated several plasma indicators in the Turkish patients with NIDDM: (i) homocysteine (Hcy) and cysteine (Cys) which contribute to increase the risk of atherosclerosis during NIDDM, (ii) glutathione (GSH) and cysteinylglycine (CysGly) resulting from GSH degradation catalyzed by γ-glutamylcysteine transferase (GGT), (iii) malonaldehyde (MDA) as a marker for lipid peroxidation, and (iv) total antioxidant status (TAS). Our main results were evaluated based on sex and diabetic status. In female patients, plasma concentrations of MDA and Hcy were significantly higher than in controls, while GSH levels were significantly lower. In males, a difference between control and diabetic groups was noticed only for Hcy, levels being also higher in patients. In the diabetic group, increase in serum glucose concentration was significantly correlated with increased GGT activity. In both controls and diabetic patients, GGT activity was correlated with a raised Cys concentration and a decreased GSH level. In both controls and diabetic patients, there were significant positive correlations between Cys and Hcy and between GSH and Hcy. We concluded that GSH and MDA levels are clinical indicators for an oxidative process linked to type 2 diabetes mellitus, especially in women. ——— NIDDM; oxidative stress; lipid peroxidation; sex © 2003 Tohoku University Medical Press

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One of the principal causes of morbidity and mortality in the individuals with diabetes mellitus is atherosclerosis and resulting cardio-vascular diseases (CVD) (Boemi et al. 1993). Type 2 diabetes mellitus with an increasing incidence world wide is characterized by an increased risk for the development of neuro-pathic, micro- and macro-vascular complications (Assmann et al. 1999). Several experimental, epidemiologic and clinical studies support the notion that oxidative stress plays a significant role in type 2 diabetes mellitus (De Zwart et al. 1999) and in the development of CVD (Jacob and Burri 1996; Garewal 1997).

The plasma thiols measurement appears to be a suitable parameter to investigate the numerous pathological diseases that are related to oxidative stress processes. Hyperhomocysteinemia has been suggested to promote the production of hydroxyl radicals, lipid peroxidation initiators, through homocysteine (Hcy) autooxidation and thiolactone formation (Stamler et al. 1993). Moreover, Hcy represents a strong and independent risk factor for CVD (Ueland et al. 1992; Boushey et al. 1995; Graham et al. 1997). The incidence of CVD increases with increasing plasma or serum Hcy concentrations (Ueland et al. 1992; Boushey et al. 1995). Furthermore, hyperhomocysteinemia has been described in diabetic patients (Munshi et al. 1996; Hoogeveen et al. 1998). Though less reactive than Hcy, cysteine (Cys) exhibits some of its chemical properties through the sulfhydryl group (Stamper and Slivka 1996). The potential noxious effects of Cys may have been overlooked since few studies analysed the relationship between Cys and CVD (Araki et al. 1989; Mansoor et al. 1995; Verhoef et al. 1996). GSH plays an important role as a scavenger of oxidant species and free radicals (Meister and Tate 1976). The precursors involved into GSH biosynthesis, i.e. Cys and γ-glutamylcysteine (γ-GluCys), and cysteinylglycine (CysGly) resulting from the reaction catalysed by γ glutamyltransferase (GGT E.C.2.3.2.2), are also of interest to evaluate the GSH homeostasis in the biological system, e.g., cells, tissues, blood or plasma (Stamler and Slivka 1996). Moreover, CysGly is a highly reactive metabolite and can produce reactive oxygen species (ROS) in presence of transition metals (Pompella 1997; Drozdz et al. 1998) thereby leading to lipid peroxidation (LPO). Serum γ -glutamylcysteine transferase (GGT) is directly involved in GSH metabolism and also modulates Cys and CysGly levels. Perry et al. (1998) suggested that an increased serum GGT level is an independent risk factor for CVD in NIDDM. Serum GGT level might be a simple and reliable marker of hepatic insulin resistance.

In the same way, the capacity of a subject to resist oxidative stress is indicated by the total antioxidant status (TAS) (Miller et al. 1993; Maxwell et al. 1997). In serum, several antioxidant compounds contribute to TAS mainly albumin, uric acid, α -tocopherol, ascorbic acid and bilirubin (Cao and Prior 1998; Diaz et al. 1998).

Determination of GSH, MDA (as a free radical-mediated LPO product) and TAS is therefore of practical and clinical importance in order to assess the implication of free radicals in type 2 diabetes. In the present study involving Turkish subjects, the main objective was to determine whether there was a link between clinical parameters and (i) the status of thiols and GGT activity in type 2 diabetes, (ii) the increase of free radical activity, measured as MDA, and (iii) the TAS in diabetic and healthy subjects.

MATERIALS AND METHODS

Subjects

Patients with type 2 diabetes mellitus (52 men and 55 women) were enrolled. All patients were taking oral antidiabetic drugs. The diagnosis of type 2 diabetes mellitus was based on the criteria of The Expert Committee on the diagnosis of diabetes mellitus (2000). Diabetic patients were recruited from those consecutive-

ly attending the Turkish Diabetes Hospital, Istanbul for their routine clinical examination (every 1–2 months) and included in the study, in agreement with Turkish local ethic committee. Non diabetic control subjects were recruited from those attending their routine health screening in the Central Laboratory of Cerrahpasa Medical Faculty (Istanbul University, Turkey). As controls, 99 healthy subjects (46 men and 53 women) were studied. No patients in the study were related. All diabetic patients had normal hepatic and endocrine functions and were relatively well controlled with glycosylated hemoglobin (HbA_{1c}) $\leq 6-7\%$ (normal range $\leq 8\%$). Tobacco and alcohol consumptions were collected by using standardised questionnaire. The patients with macro- and microangiopathic complications were excluded from this study. Determination of baseline cardiovascular risk factors included age, body mass index (BMI) (Angelico et al. 1990), alcohol consumption and smoking status.

Blood sampling

After an overnight fast, blood was drawn by venipuncture at the antecubital vein from the subjects between 8:00 and 10:00 a.m. The samples were collected in two vacuum tubes: one tube with EDTA, one tube without anticoagulant. Blood was centrifuged promptly at 1000 g for 15 minutes at +4C for plasma and serum.

Assays of biochemical parameters

Total cholesterol (TC), triglyceride (TG) and glucose were measured in serum on a Cobas-Mira analyzer (Roche Diagnostics, Meylan, France). Serum were analysed without preteatment and diluted in double-distilled water when lipid levels exceeded reference values. Inter-assay coefficients of variation of this method were 4.7%, 2.1% and 1.0%, for TG, TC and glucose, respectively. Blood glucose values were measured from venous blood serum.

GGT activity was measured in plasma

samples using a spectrophotocolorimetric assay kit (Granutest25®, Merck) on a Cobas-Mira analyser. Intra- and inter-assay coefficients of variations were 2.0% and 2.4%, respectively.

Serum TAS was determined on a Cobas-Fara analyser (Roche Diagnostics) with a "Total Antioxidant Status" kit (Randox Labs., Crumlin, UK). The assay principle relies upon the ability of antioxidants within biological fluids to quench the absorbance (measured at 600 nm) of the radical cation formed by the reaction of a chromogen (2, 2'-azino-di-[ethylbenzthiazoline sulfonate]; ABTS®) with a peroxidase and H_2O_2 (Miller et al. 1993). The system calibrator is $Trolox^{\$}$, a water-soluble vitamine E analogue. Results are expressed as μ mol Trolox/l. Intra- and inter-assay coefficients of variations were 1.9% and 3.5%, respectively.

Total plasma low molecular mass thiols (all forms including disulfides, mixed disulfides, and free thiols), i.e. Cys, CysGly, Hcy and GSH, were measured simultaneously using a high-performance liquid chromatography (HPLC) method including a precolumn derivatization (with 7-fluoro-2, 1, 3-benzoxadiazole-4-sulfonamide; ABD-F) and fluorescence detection (Salazar et al. 1999). Intra- and inter-assay coefficients of variations measured with a plasma pool was 2.2% and 7.2% for Cys, 1.7% and 3.1% for CysGly, 1.9% and 4.2% for Hcy and 7.2% and 10% for GSH at concentrations of 178.0, 27.8, 7.9 and 3.4 μ mol/liter, respectively.

The HPLC method for plasma MDA measurements (Young and Trimble 1991) consists in the following steps: (i) precipitation of proteins by phosphoric acid in order to eliminate the interference by the water-soluble substances that react with the thiobarbituric acid (TBA), (ii) formation of the TBA-MDA adduct and (iii) separation in a HPLC system with fluorescence detection. Intra- and inter-assay coefficients of variations were 5.1% and 10.5%, respectively.

Statistical analysis

Statistical analyses were performed using the BMDP® statistical software (UCLA, Los Angeles, CA, USA). Values were expressed as mean \pm s.d. Comparison between groups was performed using Student t-test or ANOVA. Ascendant stepwise multiple regression was performed to analyze interrelationships between parameters of interest (thiols, MDA and TAS levels) and the following factors: age, sex, BMI, tobacco consumption, alcohol consumption, serum cholesterol, triglycerides, apo AI, apo B and glucose concentration, HbA_{1c} level, and GGT activity. Statistical significance for all tests was accepted at $p \leq 0.05$ level.

RESULTS

The main characteristics of controls and

patients are shown in Table 1. In females, patients with NIDDM had higher TC, apo B and lower apo A1 concentrations than controls, difference for apo A1 level being only borderline significant. In males, characteristics did not differ significantly, except for glucose, between controls and patients. In female patients, plasma concentrations of MDA and Hcy were significantly higher than in controls, while GSH levels were significantly lower. In males, a difference between diabetic and control groups was noticed only for Hcy, levels being higher in type 2 diabetics.

The association between oxidative stress indicators and their clinical or biological determinants is shown in Table 2. Glucose concentration, describing the diabetic status, is significantly correlated with a reduced serum TAS. Even without significant difference of GGT

| Table 1. | Main characteristics, TAS, and, concentrations of MDA and thiol in controls (n= | |
|----------|---|--|
| | 99) and type 2 diabetic patients $(n=107)^a$ | |

| | N | Ien | Women | | |
|---|-----------------|--------------------|-----------------|--------------------|--|
| | Control (n=46) | Diabetes $(n=52)$ | Control (n=53) | Diabetes (n=55) | |
| Age (years) | 55.9 ± 14.5 | 57.8 ± 11.8 | 55.2 ± 9.1 | 57.9 ± 8.25 | |
| Body mass index (kg/m²) | 21.6 ± 3.2 | 21.8 ± 3.1 | 23.4 ± 3.5 | 23.8 ± 3.9 | |
| Smoking (pack-year) | 1.9 ± 1.2 | 2.1 ± 0.9 | 1.1 ± 0.8 | 0.9 ± 0.7 | |
| Alcohol (g/week) | 0.07 ± 0.26 | 0.02 ± 0.15 | _ | _ | |
| Serum glucose (mmol/liter) | 3.35 ± 0.56 | $9.07 \pm 3.83***$ | 3.54 ± 0.52 | $9.08 \pm 3.80***$ | |
| $\mathrm{HbA}_{\mathrm{1c}}$ (%) | ND | 6.7 ± 1.9 | ND | 6.9 ± 2.2 | |
| Serum triglycerides (mmol/liter) ^b | 1.48 ± 0.55 | 2.18 ± 2.12 | 1.65 ± 0.67 | 1.70 ± 0.95 | |
| Serum cholesterol (mmol/liter) | 4.89 ± 1.46 | 5.17 ± 1.05 | 5.30 ± 1.10 | $5.86 \pm 1.04*$ | |
| Apo A1 (g/liter) | 1.28 ± 0.25 | 1.27 ± 0.24 | 1.54 ± 0.29 | 1.44 ± 0.22 | |
| Apo B (g/liter) | 1.11 ± 0.26 | 1.11 ± 0.31 | 1.07 ± 0.25 | $1.18 \pm 0.25*$ | |
| GGT activity (U/liter) ^b | 20.9 ± 2.8 | 25.0 ± 3.1 | 20.0 ± 1.7 | 21.9 ± 2.4 | |
| MDA (µmol/liter) | 0.36 ± 0.15 | 0.43 ± 0.19 | 0.37 ± 0.12 | $0.47 \pm 0.23**$ | |
| TAS (µmol/l equiv Trolox®) | 1.40 ± 0.12 | 1.43 ± 0.14 | 1.39 ± 0.14 | 1.42 ± 0.12 | |
| Plasma cysteine (µmol/liter) | 279 ± 42 | 291 ± 53 | 269 ± 57 | 282 ± 57 | |
| Plasma cysteinylglycine (µmol/liter) | 41.1 ± 7.7 | 40.6 ± 13.5 | 37.0 ± 8.3 | 37.9 ± 10.8 | |
| Plasma homocysteine (µmol/liter) | 11.2 ± 3.6 | $14.6 \pm 5.7*$ | 9.4 ± 2.7 | $11.4 \pm 3.7**$ | |
| Plasma glutathione (μ mol/liter) | 7.7 ± 2.1 | 7.3 ± 3.1 | 7.8 ± 2.7 | $6.4 \pm 3.4*$ | |

^aValues are means ± s.D.

^bTest on log-transformed values.

^{*} $p \le 0.05$, ** $p \le 0.01$, *** $p \le 0.001$: Student's t-test between controls and patients or χ^2 test ND, not determined.

Table 2. Determinants of TAS, MDA and thiol levels in (A) controls (n=99) and (B) type 2 diabetic patients (n=107) (both males and females)^a

A (Controls)

| | TAS (µmol/l) | MDA (µmol/l) | Cys (µmol/l) | CysGly (µmol/l) | Hcy (µmol/l) | GSH (µmol/l) |
|----------------------------|-----------------|-----------------|-----------------|--------------------|-----------------|-----------------|
| Age (years) | _ | _ | 1.27(0.49)** | _ | _ | 0.067(0.028)* |
| Sex | _ | _ | _ | _ | -3.11(1.16)** | _ |
| $BMI (kg/m^2)$ | _ | _ | _ | _ | _ | _ |
| Serum cholesterol (mmol/l) | _ | _ | 14.5(4.9)** | _ | _ | _ |
| GGT activity (U/l)b | _ | _ | 47.9(22.8)* | _ | _ | $-2.67(1.25)^*$ |
| Intercept | _ | _ | 70.8 | _ | 17.66 | 7.72 |
| \mathbb{R}^2 | _ | _ | 0.238 | _ | 0.095 | 0.161 |

B (Diabetic patients)

| | TAS (µmol/l) | MDA (µmol/l) | Cys (µmol/l) | CysGly (µmol/l) | Hcy (µmol/l) | GSH (µmol/l) |
|---------------------------|------------------|-----------------|-----------------|--------------------|-----------------|-----------------|
| Age (years) | _ | _ | 2.54(0.62)*** | _ | 0.134(0.036)*** | _ |
| Sex | _ | _ | _ | _ | -2.13(0.68)** | _ |
| $BMI (kg/m^2)$ | _ | 0.015(0.006)* | _ | _ | _ | _ |
| Serum glucose (mmol/l) | 0.0093(0.0036)** | _ | _ | _ | _ | _ |
| GGT activity (U/l)b | 0.178(0.053)*** | _ | 60.6(21.8)** | _ | _ | 4.61(1.31)*** |
| Intercept | 1.28 | 0.876 | 56.4 | _ | 5.60 | 12.66 |
| \mathbb{R}^2 | 0.175 | 0.077 | 0.243 | _ | 0.224 | 0.134 |

^aRegression coefficient (standard error).

activity between the control and diabetic groups, in both controls and diabetic patients GGT activity is related with a raised Cys concentration and a decreased GSH level. Nevertheless, no potential activity of GGT on GSH was detected after the blood uptake: GSH concentration did not change during sample treatment in the range of GGT activities met in the present study (data not shown).

The relationships between oxidative stress indicators are summarized in Table 3. In both controls and diabetic patients, there are significant positive correlations between Cys and Hcy and between GSH and Hcy.

DISCUSSION

Patients with NIDDM have increased mortality and morbidity compared with non-diabetics and are more likely to develop CVD (Boemi et al. 1993). The high risk for vascular disease can only partly be explained by the traditional risk factors for the general population such as smoking, hypertension and raised cholesterol. It has been proposed that oxidative stress may be associated with the pathogenesis of the complications of NIDDM (De Zwart et al. 1999), particularly CVD (Jacob and Burri 1996; Garewal 1997).

One issue that has not previously been addressed is the interrelationship between thiols

^bLog-transformed values.

NS not significant, * $p \le 0.05$, ** $p \le 0.01$, *** $p \le 0.001$.

TAS, Total antioxidant status; MDA, Malonaldehyde; GSH, Glutathione.

Table 3. Correlation coefficients between TAS, MDA and thiol levels in (A) controls (n=99) and (B) type 2 diabetic patients (n=107) (both males and females) adjusted for age and sex

| A | (Controls) | ١ |
|---|------------|---|
| | | |

| | TAS | MDA | Cys | CysGly | Нсу |
|--------|-----------------|--------|-----------------|-----------------|-------------|
| MDA | -0.012 | _ | _ | _ | _ |
| Cys | -0.185 | 0.034 | _ | _ | _ |
| CysGly | $0.241^{\rm b}$ | 0.143 | $0.350^{\rm c}$ | _ | _ |
| Hcy | 0.152 | -0.143 | 0.332^{c} | $0.272^{\rm b}$ | _ |
| GSH | 0.085 | -0.156 | 0.027 | 0.157 | 0.298^{b} |

B (Diabetic patients)

| | TAS | MDA | Cys | CysGly | Нсу |
|--------|-----------------|-------|------------------|-----------------|-----------------|
| MDA | -0.004 | _ | _ | _ | _ |
| Cys | 0.108 | 0.078 | _ | _ | _ |
| CysGly | 0.258^{b} | 0.159 | -0.101 | _ | _ |
| Hcy | $0.293^{\rm b}$ | 0.171 | $0.275^{\rm b}$ | $0.431^{\rm d}$ | _ |
| GSH | -0.201^{a} | 0.092 | $-0.266^{\rm b}$ | $0.531^{\rm d}$ | $0.291^{\rm b}$ |

 $^{a}p \le 0.10, ^{b}p \le 0.05, ^{c}p \le 0.01, ^{d}p \le 0.001$

(i.e., Cys, CysGly, Hcy and GSH) status and type 2 diabetes. According to a previous study (Chico et al. 1998), diabetic patients had higher Hcy levels than control subjects. Moreover, the observed sex-related difference (Hcy concentrations lower in female than male patients and controls) implies that Hcy metabolism may not differ between diabetic patients and control subjects. Das et al. (1999) suggested that NIDDM may strongly promote oxidative stress and enhance CVD in patients who have coexistent hyperhomocysteinemia. Despite no significant variation of Cys and CysGly concentrations was observed in our study, the increase of oxidative stress is confirmed, for women, in diabetic patients with lower GSH than in controls (Sundaram et al. 1996) and with a GGT activity related to oxidative stress in type 2 diabetes (Perry et al. 1998). The relationship between GGT activities and its pro-oxidant effect has been demonstrated in vitro (Drozdz et al. 1998; Enoiu et al. 2000) and in clinical studies by the increase in lipid peroxidation (Pompella

1997; Perry et al. 1998). Our findings were in agreement with Sundaram et al. (1996) stressing to an increase in lipid peroxidation in patients with type 2 diabetes mellitus. Using the ferrous-oxidation of xylenol orange assay to determine lipid hydroperoxides, Nourooz-Zadeh et al. (1997) suggested that changes in oxidative stress were related to the underlying metabolic abnormalities in type 2 diabetes mellitus rather than to the appearance of complications. This is supported by the finding of raised LPO products in patients with impaired glucose tolerance, even before the onset of frank diabetes (Vijayalingam et al. 1996). In our study, a raised LPO was demonstrated in female diabetic patients with higher MDA concentrations than in controls. The higher level of oxidative stress in women might be because of their higher BMI levels. The average plasma concentrations of MDA measured in our healthy adults were similar to those found by Carbonneau et al. (1991), and higher values observed in diabetic patients were described by Langley et

al. (1993) in ill subjects concerned by such oxidative stress related pathologies. Moreover, in healthy individuals, oxidative damage was reported to be more extensive in women than in men in a study performed by Block et al. (2002), whereas Ide et al. (2002) found enhanced oxidative stress in men compared to women.

The TAS should reflect the ability of an individual to resist oxidative stress. Low serum antioxidative activity in NIDDM may be related to an increased tendency to LPO. According to our results, there was no significant difference in serum TAS between diabetic patients and control subjects, which is in contrast to other reports showing a decreased antioxidative activity in diabetic patients (Ceriello et al. 1997; Maxwell et al. 1997) but in concordance with Leinonen et al. (1998). However, the relevance of the Randox-TAS® has been recently discussed in comparison with other "antioxidant assays", i.e., the oxygen radical absorbance capacity (ORAC) and the ferric reducing ability of plasma (FRAP) assays (Cao and Prior 1998): a low correlation was pointed out between TAS and the two other methods. Nevertheless, TAS is also an easier approach than specific methods such as hydroxylation of aspirin (generation of hydroxyl radical) already used in NIDDM (Ghiselli et al. 1992). Several studies have consistently demonstrated deficiency in individual antioxidants in NIDDM patients (Sundaram et al. 1996; Vijayalingam et al. 1996; Ceriello et al. 1997; Nourooz-Zadeh et al. 1997) with reduced concentrations of GSH, vitamin C and the antioxidant enzymes, i.e., superoxide dismutase and catalase, as well as a decrease in total radical trapping antioxidant parameter, suggesting a reduced total antioxidant defence. Overall, these studies assume that oxidative stress and impaired antioxidant defence is a feature of type 2 diabetes mellitus that is present early in the disease and may contribute to its progression and even to the development of complications.

In the present study, we have confirmed that type 2 diabetic women patients developed an oxidative stress by the decline in antioxidant defense including a lowered GSH (as a major antioxidant) level and an increased MDA (as a LPO product) concentration. These findings are of particular interest in view of the fact that women might be at a greater risk of diabetes than men. The increased oxidative damage that we have pointed out may therefore predispose to the development of CVD. Profiling the oxidative stress parameters status in such a pathology may contribute to protection from CVD in the female population.

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