

Insights into the binding mode of new N-substituted pyrazoline derivatives to MAO-A: docking and quantum chemical calculations

Safiye Sağ Erdem · Seyhan Türkkkan ·
Kemal Yelekçi · Nesrin Gökhan-Kelekçi

Received: 13 October 2012 / Accepted: 3 December 2012 / Published online: 16 December 2012
© Springer-Verlag Wien 2012

Abstract The binding modes of four N-substituted pyrazoline derivatives as novel MAO-A inhibitory agents were investigated using docking and quantum chemical molecular modelling tools.

Keywords N-substituted pyrazolines · Docking · PM6

Introduction

Monoamine oxidase (MAO) is a flavoprotein located in the outer membrane of the mitochondria that contains a covalently bound flavin adenine dinucleotide (FAD) as a coenzyme and that has considerable physiological and pharmacological interest due to its central role in the metabolism of monoamine neurotransmitters. MAO exists in two isoforms, MAO-A and MAO-B, which share approximately 70 % sequence identity on the amino acid levels and differ in their substrate specificity, susceptibility to specific inhibitors, and three-dimensional structure (Binda et al. 2004; Edmondson et al. 2004). Since they metabolize the principal biogenic amines, MAO-A and MAO-B play an important role in the regulation of their

concentrations mainly in the central nervous system, where abnormal values have been involved in psychiatric and neurodegenerative disorders such as, depression, Alzheimer's disease, and Parkinson's disease (Shih 2004; Riederer et al. 2004). Selective inhibition of MAO-A results in elevated noradrenaline and serotonin concentrations, thus gradually improving the symptoms of depression. On the other hand, inhibition of MAO-B is a crucial strategy for treatment of Parkinson disease (Li et al. 2006). Indeed, treatment of pre-Parkinson's patients with selective MAO-B inhibitors has been shown to be effective in reducing the development of this neurodegeneration. All these findings support the clinical importance of MAO inhibitors in the treatment of several neurological and psychiatric disorders. In the light of these knowledge and the previous findings (Jayaprakash et al. 2008; Gökhan-Kelekçi et al. 2007), we synthesized N-substituted pyrazoline derivatives (Fig. 1) as novel potential MAO inhibitory agents which were found to be selective to MAO-A. Continuing our efforts in computational studies (Erdem and Yelekçi 2001; Toprakçi and Yelekçi 2005; Erdem et al. 2006; Akyüz et al. 2007; Yelekçi et al. 2007; Erdem and Büyükmenekşe 2011), we aimed to present molecular insights into the binding modes of these compounds in the active site of MAO-A through the use of molecular modelling tools. Ultimate aim of this study is to contribute to the design and development of more effective and selective inhibitors than the ones presently involved into clinical studies such as rasagiline and selegiline.

Materials and methods

The MAO-A crystal structure was obtained from protein data bank, code 2Z5X (Son et al. 2008). The ADT (Auto Dock Tools) package (Michel and Saner 1999) was

S. S. Erdem (✉) · S. Türkkkan
Department of Chemistry, Faculty of Arts and Sciences,
Marmara University, 34722 Göztepe, Istanbul, Turkey
e-mail: erdem@marmara.edu.tr

K. Yelekçi
Bioinformatics and Genetics Department, Faculty of
Engineering and Natural Sciences, Kadir Has University,
34083 Fatih, Istanbul, Turkey

N. Gökhan-Kelekçi
Department of Pharmaceutical Chemistry, Faculty of Pharmacy,
Hacettepe University, 06100 Sıhhiye, Ankara, Turkey

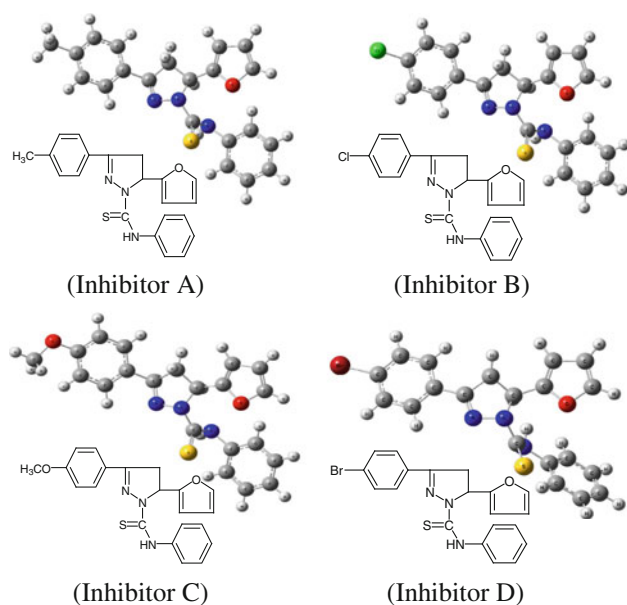


Fig. 1 Structures of the inhibitors studied

employed to generate the docking input files of the protein and four inhibitors. Hydrogen atoms were added to the crystal structure, and partial atomic charges were calculated via the Gasteiger–Marsili method (Gasteiger and Marsili 1980).

Prior to docking, the system was subjected to 10 ns of molecular dynamic simulation at 310 K using NAMD v2.6. (Phillips et al. 2005). The docking procedure utilized in previous studies (Toprakçi and Yelekçi 2005; Yelekçi et al. 2007) was used. However, some differences in the procedure are as follows: AutoDock 4.2 (Morris et al. 2009) was employed to perform the docking simulation using a Lamarckian genetic algorithm (Morris et al. 1998; Huey et al. 2007). A grid of 70, 70, 70 points in x , y , and z directions was built on the center of mass of the N5 atom of the flavin. The structure with the most favorable free energy of binding was selected and analyzed using the Accelrys Software 3.1 (Discovery Studio Modeling Environment 2011).

The resultant docked structures were analyzed using the Gaussview 5.0 program. 23 amino acids in the active site surrounding the inhibitor were selected and the remaining amino acids were removed from the structure. 14 water molecules were then inserted into the truncated structure with the same coordinates as in the X-ray structure. The resultant structure consisted of about 550 atoms. Three methyl carbons of FAD and two backbone carbons of side chains were frozen to prevent unnatural changes in the structure prior to optimization. Geometry optimization was performed employing semi-empirical PM6 method (Rezác et al. 2009) in Gaussian 09 (Frisch et al. 2009).

Results and discussion

The inhibition constants, K_i , calculated from docking are 6.9, 8.5, 5.4, and 4.3 nM for inhibitors A, B, C, and D, respectively, and are in good agreement with the experimental results (manuscript in preparation). Tight binding interactions with MAO-A enzyme are expected since these compounds have highly potent K_i values. All four compounds bind to the *re*-face of FAD in the active site of MAO-A (Fig. 2). A common feature of their binding mode is the packing of the furan moiety between the phenolic side chains of Tyr407 and Tyr444 so that binding can be enhanced through a favorable π – π stacking interaction. The distances between furan oxygen and Tyr444 are 4.00, 3.45, 3.87, and 3.66 Å for A, B, C, and D, respectively. Corresponding distances with Tyr407 are 4.02, 4.38, 4.14, and 3.73 Å for A, B, C, and D, respectively. The *p*-substituents of the phenyl groups are situated near Lys305, facilitating attractive interactions in C and D (2.14 and 2.73 Å, respectively). Another common feature is the extension of the phenyl substituent of the thiosemicarbazide moiety towards Ileu180 and Asn181. NH hydrogen in thiosemicarbazide moiety acts as a hydrogen bond donor to the nearby side chains Ile180 in B (2.27 Å) and Asn181 in D (2.07 Å).

Moreover, additional hydrogen-bonding interactions are observed between hydroxyl hydrogen of Tyr444 and thiosemicarbazide sulfur atoms for A and C (2.46 and 2.59 Å, respectively).

Docking theories are known to be less sensitive than quantum mechanical methods in predicting electronic interactions. As a quantum mechanical semi-empirical method, the recently developed PM6 method (Rezác et al. 2009) was employed here, since it achieved major improvements in accuracy for the interaction energies of biologically relevant, non-covalently bound systems, with empirical corrections for dispersion and hydrogen-bonding interactions (Stewart 2007). The superposition of PM6-optimized structures with those of docked orientations is shown in Fig. 2. A key advantage of PM6 calculations is that they allow us to observe the interactions between water and active site protein residues as well as water and each inhibitor. Such information cannot be gained by docking calculations since water molecules are excluded from the enzyme.

Several discrepancies are noticeable in the PM6-optimized structures, mainly as the result of the interactions of water molecules. All four inhibitors show H-bonding type attractions between the sulfur atom in thiosemicarbazide and the hydrogen of the nearby water molecule. The interaction distances are predicted to be 2.75, 2.63, 2.65, and 2.68 Å for inhibitors A, B, C, and D, respectively. In C, *p*-OCH₃ hydrogen atoms exhibit non-bonded interactions

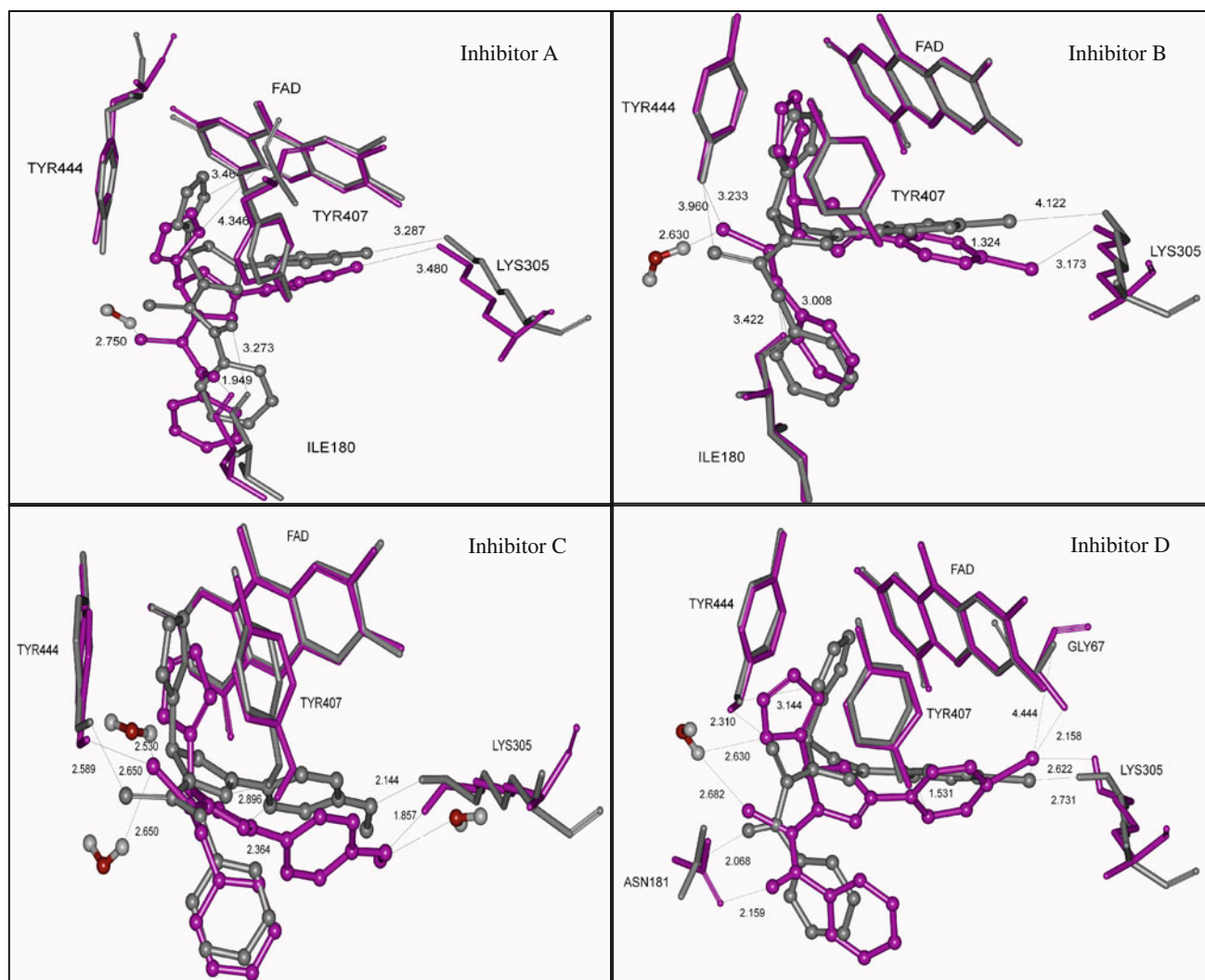


Fig. 2 Binding orientation of inhibitors in MAO-A (grey: docking, purple: PM6). Remaining side chains are not shown for clarity

with the oxygen of two different water molecules having distances of 2.53 and 2.15 Å.

The orientation of the furan moiety is slightly altered after PM6 optimization. In C, it adopts a nearly perpendicular conformation with respect to Tyr407 and Tyr4444, contrary to the alignment in A, B, and D in which it aligns in a parallel manner with the tyrosines. Only in inhibitor D does the furan oxygen point away from FAD and creates a H-bond with the nearby water molecule which is also H-bonded to the thiosemicarbazide sulfur atom. As a result, the furan ring is pushed slightly out of the aromatic cage facilitating an additional H-bond with Tyr444 hydroxyl hydrogen (2.31 Å), which is a very weak interaction in the docked structure (3.13 Å).

Similarly, some other binding modes appear to be stronger, which is evident from the shorter interaction distances in PM6-optimized structures. The H-bond

distance to Lys305 shortens from 2.14 to 1.86 Å in C, and from 2.73 to 2.62 Å in D.

Conclusion

Several discrepancies were observed between the docked pose of the compounds and the optimized pose obtained from the quantum chemical PM6 method, which presents more realistic interactions. A general feature of the binding mode in both methods is that the furan moiety of the inhibitors aligns in between the aromatic cage tyrosine residues (Tyr407 and Tyr444) and interacts either in π - π stacking interactions or H-bonding interactions with the hydroxyl group of tyrosines. Interactions with active site water molecules provided additional insights into the binding modes of the inhibitors studied. Considering the

calculated K_i values (4.28–8.51 nM) and strong binding interactions, these compounds appear to be promising MAO-A inhibitors.

Acknowledgments This work was supported by TUBITAK, project no. 108T232, and Marmara University Scientific Research Projects Commission (BAPKO), project no. FEN-D-130612-0243. The authors thank Dr. D. Akten for her technical assistance for the MD simulation and Dr. V.E. Atalay for the Gaussian calculations.

References

- Akyüz M, Erdem SS, Edmondson DE (2007) Aromatic cage in the active site of monoamine oxidase B: effect on the structural and electronic properties of bound benzylamine and *p*-nitrobenzylamine. *J Neural Transm* 114:693–698
- Binda C, Huba'lek F, Li M, DE Edmondson, Mattevi A (2004) Crystal structure of human monoamine oxidase B, a drug target enzyme monotonically inserted into the mitochondrial outer membrane. *FEBS Lett* 564:225–228
- Discovery Studio Modeling Environment (2011) Release 3.1. Accelrys Software Inc., San Diego
- Edmondson DE, Mattevi A, Binda C, Li M, Huba'lek F (2004) Structure and mechanism of monoamine oxidase. *Curr Med Chem* 11:1983–1993
- Erdem SS, Büyükmekşe B (2011) Computational investigation on the structure-activity relationship of the biradical mechanism for monoamine oxidase. *J Neural Transm* 118:1021–1029
- Erdem SS, Yelekçi K (2001) Computer modeling of oxygen containing heptylamines as monoamine oxidase inactivators. *J Mol Struct Theochem* 572:97–106
- Erdem SS, Karahan Ö, Yıldız İ, Yelekçi K (2006) A computational study on the amine-oxidation mechanism of monoamine oxidase: insight into the polar nucleophilic mechanism. *Org Biomol Chem* 4:646–658
- Frisch MJ et al (2009) Gaussian 09, Revision A1. Gaussian, Inc., Wallingford CT
- Gasteiger J, Marsili M (1980) Iterative partial equalization of orbital electronegativity—a rapid access to atomic charges. *Tetrahedron* 36:3219–3228
- Gökhan-Kelekçi N, Küpeli E, Salgın U, Özgen Ö, Uçar G, Yeşilada E, Kendi E, Yeşilada A, Bilgin AA (2007) A new therapeutic approach in Alzheimer disease: some novel pyrazole derivatives as dual MAO-B inhibitors and antiinflammatory analgesics. *Bioorg Med Chem* 15:5775–5786
- Huey R, Morris GM, Olson AJ, Goodsell DS (2007) A semi-empirical free energy force field with charge-based desolvation. *J Comput Chem* 28:1145–1152
- Jayaprakash V, Sinha BN, Uçar G, Ercan A (2008) Pyrazoline-based mycobactin analogues as MAO-inhibitors. *Bioorg Med Chem Lett* 18:6362–6368
- Li M, Binda C, Mattevi A, Edmondson DE (2006) Functional role of the “aromatic cage” in human monoamine oxidase b: structures and catalytic properties of Tyr435 mutant proteins. *Biochemistry* 45:4775–4784
- Michel F, Saner P (1999) Python: a programming language for software integration and development. *J Mol Graph Model* 17:57–61
- Morris GM, Goodsell DS, Halliday RS, Huey R, Hart WE, Belew RK, Olsan AJ (1998) Automated docking using a Lamarckian genetic algorithm and empirical binding free energy function. *J Comp Chem* 19:1639–1662
- Morris GM, Huey R, Lindstrom W, Sanner MF, Belew RK, Goodsell DS, Olson AJ (2009) Autodock4 and AutoDockTools4: automated docking with selective receptor flexibility. *J Comput Chem* 16:2785–2791
- Phillips JC, Braun R, Wang W, Gumbart J, Tajkhorshid E, Villa E, Chipot C, Skeel RD, Kale L, Schulten K (2005) Scalable molecular dynamics with NAMD. *J Comput Chem* 26:1781–1802
- Rezac J, Fanfrlik J, Salahub D, Hobza P (2009) Semiempirical quantum chemical PM6 method augmented by dispersion and h-bonding correction terms reliably describes various types of noncovalent complexes. *J Chem Theory Comput* 5:1749–1760
- Riederer P, Lachenmayer L, Laux G (2004) Clinical applications of MAO inhibitors. *Curr Med Chem* 11:2033–2043
- Shih JC (2004) Cloning, after cloning, knock-out mice and physiological functions of MAO A and B. *Neurotoxicology* 25:21–30
- Son Y, Ma J, Kondou Y, Yoshimura M, Yamashita E, Tsukihara T (2008) Structure of human monoamine oxidase A at 2.2-Å resolution: the control of opening the entry for substrates/inhibitors. *Proc Natl Acad Sci USA* 105:5739–5744
- Stewart JJP (2007) Optimization of parameters for semiempirical methods V: modification of NDDO approximations and application to 70 elements. *J Mol Mod* 13:1173–1213
- Toprakçi M, Yelekçi K (2005) Docking studies on monoamine oxidase-B inhibitors: estimation of inhibition constants (K_i) of a series of experimentally tested compounds. *Bioorg Med Chem Lett* 15:4438–4446
- Yelekçi K, Karahan Ö, Toprakçi M (2007) Docking of novel reversible monoamine oxidase-B inhibitors: efficient prediction of ligand binding sites and estimation of inhibitors thermodynamic properties. *J Neural Transm* 114:725–732